Fiscal Year:	FY 2015	Task Last Updated:	FY 07/07/2015
PI Name:	Mellor, Liliana F. Ph.D.	Å	
Project Title:	Induction of Early Stages of Osteoarthritis	After Exposure to Microgravity	
Division Name:	Human Research		
Program/Discipline:	NSBRI		
Program/Discipline Element/Subdiscipline:	NSBRIMusculoskeletal Alterations Tean	n	
Joint Agency Name:		TechPort:	No
Human Research Program Elements:	(1) HHC :Human Health Countermeasures		
Human Research Program Risks:	 Bone Fracture: Risk of Bone Fracture Osteo: Risk Of Early Onset Osteoporos 	due to Spaceflight-induced Chang sis Due To Spaceflight	es to Bone
Space Biology Element:	None		
Space Biology Cross-Element Discipline:	None		
Space Biology Special Category:	None		
PI Email:	lfmellor@ncsu.edu	Fax:	FY
PI Organization Type:	UNIVERSITY	Phone:	208-426-2238
Organization Name:	North Carolina State University		
PI Address 1:	Department of Biomedical Engineering		
PI Address 2:	Cell Mechanics Laboratory		
PI Web Page:			
City:	Raleigh	State:	NC
Zip Code:	27695-7115	Congressional District:	4
Comments:	NOTE: formerly at Boise State University	until fall 2013 (Ed., Jan 2014)	
Project Type:	Ground	Solicitation / Funding Source:	2011 NSBRI-RFA-11-01 Postdoctoral Fellowships
Start Date:	11/01/2011	End Date:	02/28/2015
No. of Post Docs:	1	No. of PhD Degrees:	0
No. of PhD Candidates:	0	No. of Master' Degrees:	0
No. of Master's Candidates:	0	No. of Bachelor's Degrees:	0
No. of Bachelor's Candidates:	0	Monitoring Center:	NSBRI
Contact Monitor:		Contact Phone:	
Contact Email:			
Flight Program:			
Flight Assignment:	NOTE: End date change to 2/28/2015 per NSBRI (Ed., 12/2/14) NOTE: End date is now 11/30/2014 with PostDoc change in institution, per NSBRI (Ed., 1/15/14) NOTE: End date changed to 11/30/2013 per NSBRI (Ed., 10/24/13)		
Key Personnel Changes/Previous PI:			
COI Name (Institution):	Loboa, Elizabeth (MENTOR/North Car	olina State University)	
Grant/Contract No.:	NCC 9-58-PF02601		
Performance Goal No.:			
Performance Goal Text:			

Task Description:	POSTDOCTORAL FELLOWSHIP Little is known about the effects of spaceflight on articular cartilage health. Unlike bone and muscle, cartilage lacks the ability to regenerate. Once a catabolic cascade is triggered, it usually results in osteoarthritis. Some studies have shown degradation of the articular cartilage in response to unloading and prolonged bedrest. However, the underlying molecular mechanisms of articular cartilage degradation in response to unloading are still elusive. The overall goal of my research is to understand and elucidate key molecular mechanisms involved in response of cartilage to changes in gravitational forces. The majority of my focus is at the cell level, with analyses of potential approaches to translate these changes to tissue and joint level. To begin to address this problem, I investigated the effects of simulated microgravity on chondrocytes using the rotating wall vessel (RWV) bioraector. Also, in collaboration with Dr. Jeff Willey, I studied the combined effects of radiation and simulated microgravity on articular cartilage health. I found that chondrocytes exposed to simulated microgravity undergo morphological rearrangement of the actin cytoskeleton. This change was not observed in irradiated cells; however, when cells were exposed to both radiation and simulated microgravity express more collagen I and tegs: confirmed that cells exposed to both radiation and simulated microgravity express more collagen I and tegs: colfared that cells exposed to both radiation and simulated microgravity express more collagen I and tegs: colfared that cells exposed to both radiation. Sure regular of Wnt signaling that has been shown to protect against radiation, but not radiation and microgravity. If chondrocytes is normal gravity respond to radiation by producing "radioprotectant" genes, simulated microgravity. If chondrocytes in normal gravity respond to radiation by producing "radioprotectant" genes, simulated microgravity and infersing a combined synergistie, degradatory effect of simulated m	
Rationale for HRP Directed Research:		
Research Impact/Earth Benefits:	Astronauts are exposed to prolonged periods of microgravity during spaceflight. Lack of gravity is known to negatively affect the musculoskeletal system, mainly bone and skeletal muscle, which are constantly exposed to mechanical loading on Earth. However, another important component of the musculoskeletal system, the synovial joint, has not been fully investigated in space-like conditions. Unlike bone and muscle, the articular cartilage of the synovial joint has limited regenerative capacity, and has a relatively slow turn over compared to the fast remodeling of bone. Cartilage degradation leads to a severe disease known as osteoarthritis (OA). OA is the leading cause of disability in the US, and one of the few chronic diseases of aging without a cure. Little is known about the effects of microgravity on the synovial joint and changes in cartilage homeostasis. Bone remodeling occurs much faster. This is the first study to use an in vitro and ex vivo approach to understand the effects of reduced gravity on cartilage homeostasis and molecular pathways responsive to changes in gravitational forces. Understanding changes in cartilage homeostasis and molecular pathways in reduced gravity conditions, will help us elucidate the mechanisms involved in disuse OA here on Earth, and develop novel therapeutic targets.	
Task Progress:	This is the first study to show cartilage degradation at both the cell and tissue level in response to simulated microgravity via changes in gene expression, protein expression, and GAG (glycosaminoglycan) content. Although it appears that cartilage degradation in response to microgravity may take longer than bone density loss and skeletal muscle atrophy, cartilage tissue does not regenerate. Loss or degeneration of articular cartilage results in painful and debilitating joint disease that can limit mobility of the affected joint. We also targeted a potential signaling pathway that responds to mechanical unloading in chondrocytes. Active canonical Wnt signaling is essential in bone to maintain a balance between osteoclast and osteoblast activity. Inhibition of Wnt signaling has been correlated with bone loss. We detected inhibition of Wnt signaling in response to simulated microgravity in chondrocytes by up-regulation of sclerostin and other Wnt inhibitors. However, in cartilage, Wnt inhibition has been reported to be a chondro-protective mechanism to prevent further cartilage degradation, and canonical Wnt signaling has been associated with cartilage degradation and OA. We showed in our chondrocyte cell line that up-regulation of catabolic genes in response to simulated microgravity was similar to that of IL-1ß treatment. However, when recombinant sclerostin was added to IL-1ß treated cells, it decreased MMP (matrix metalloproteinase) expression as previously suggested, providing supportive evidence of the chondroprotective role of sclerostin in cartilage tissue. Therefore, it appears that sclerostin has opposite roles in these two adjacent tissues, and therapeutic targets using a sclerostin antibody to reduce bone density loss in space and osteoporosis need to be further evaluated given the potential of sclerostin inhibition resulting in the progression of OA. Funding for this project has helped provide new evidence for another tissue potentially affected by lack of gravity. Osteoarthritis and joint degradati	
Bibliography Type:	Description: (Last Updated: 11/12/2020)	
Articles in Peer-reviewed Journals	Mellor L, Mohiti-Asli M, Williams J, Kannan A, Dent MR, Guilak F, Loboa E. "Extracellular calcium modulates chondrogenic and osteogenic differentiation of human derived adipose stem cells: A novel approach for osteochondral tissue engineering using a single stem cell source." Tissue Eng Part A. 2015 Jun 2. [Epub ahead of print] PubMed <u>PMID: 26035347</u> ; <u>http://dx.doi.org/10.1089/ten.TEA.2014.0572</u> , Jun-2015	

Articles in Peer-reviewed Journals	Mellor LF, Baker TL, Brown RJ, Catlin LW, Oxford JT. "Optimal 3D culture of primary articular chondrocytes for use in the rotating wall vessel bioreactor." Aviation, Space, and Environmental Medicine. 2014 Aug;85(8):798-804. PubMed <u>PMID: 25199120</u> ; PubMed Central <u>PMCID: PMC4207436</u> ; <u>http://dx.doi.org/10.3357/ASEM.3905.2014</u> , Aug-2014
Articles in Peer-reviewed Journals	Mellor LF, Steward AJ, Nordberg RC, Taylor MA, Loboa EG. "Comparison of simulated microgravity and hydrostatic pressure for chondrogenesis of hASC." Aerosp Med Hum Perform. 2017 Apr;88(4):377-84. https://doi.org/10.3357/AMHP.4743.2017; PubMed PMID: 28518000, Apr-2017
Articles in Peer-reviewed Journals	Mellor LF, Nordberg RC, Huebner P, Mohiti-Asli M, Taylor MA, Efird W, Oxford JT, Spang JT, Shirwaiker RA, Loboa EG. "Investigation of multiphasic 3D-bioplotted scaffolds for site-specific chondrogenic and osteogenic differentiation of human adipose-derived stem cells for osteochondral tissue engineering applications." J Biomed Mater Res B Appl Biomater. 2020 Jul;108(5):2017-30. Epub 2019 Dec 27. <u>https://doi.org/10.1002/jbm.b.34542</u> ; <u>PMID: 31880408</u> ; <u>PMCID: PMC7217039</u> , Jul-2020
Articles in Peer-reviewed Journals	Nordberg RC, Mellor LF, Krause AR, Donahue HJ, Loboa EG. "LRP receptors in chondrocytes are modulated by simulated microgravity and cyclic hydrostatic pressure." PLoS One. 2019 Oct 4;14(10):e0223245. https://doi.org/10.1371/journal.pone.0223245 ; PMID: 31584963; PMCID: PMC6777824, Oct-2019
Awards	Mellor LF. "Career advancement award, National Space Biomedical Research Institute, December 2014." Dec-2014
Awards	Mellor LF. "Fellow Travel Award, Biomedical Engineering Society (BMES)-CMBE annual meeting, January 2015." Jan-2015
Awards	Mellor LF. "Poster competition award, 3rd place Postdoctoral Research Symposium at NC State University, May 2014." May-2014
Awards	Mellor LF. "Professional Development Award, NCSU (NC State University) Postdoctoral Association, September 2014." Sep-2014